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## Ramos cell line

Catalog Number: PC-030 (For Research Use Only)

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### Product Overview

The Ramos cell line is a human B lymphocyte cell line derived from Burkitt's lymphoma. It is widely used as a model for B cell receptor (BCR) signaling, immunology, and lymphoma biology. Ramos cells are particularly valuable for studying antigen receptor activation, apoptosis, and signal transduction in B cells.

### Materials provided

One vial of  $2 \times 10^6$  cells in Freezing Media. **IMPORTANT:** store the frozen cells in liquid nitrogen until you are ready to thaw and propagate them.

### Handling cells upon arrival

It is strongly recommended that you propagate the cells by following instructions as soon as possible upon arrival.

### Required Cell Culture Media

- **Complete Growth Media**  
In 450mL of RPMI-1640, add 50mL FBS (10% final) and 5mL Penicillin/Streptomycin (1% final).
- **Freezing Media**  
Add 10% DMSO (final) to 70% Complete Growth Media and 20% FBS. Make fresh each time.

### Materials required but not provided

(Can be substituted with a comparable third-party product)

- RPMI-1640 Medium -- Hyclone P/N SH30027.01
- Fetal Bovine Serum (FBS) -- Fisherbrand P/N 03-600-511
- Penicillin/Streptomycin -- Hyclone P/N SV30010
- Trypsin -- Hyclone P/N SH30236.02
- Phosphate-buffered saline (PBS) -- Cellgro P/N 21-040-CV
- DMSO -- Sigma P/N D8418

### Initial Culture Procedure

1. Quickly thaw cells in a 37°C water bath with careful agitation. Remove from the bath as soon as the vial is thawed, and decontaminate by dipping in or spraying with 70% ethanol.
2. Transfer cells to a 100mm<sup>2</sup> dish (or T25cm<sup>2</sup> flask) containing 10-12ml of **Complete Growth Media**.
3. Gently rock the flask to ensure the cells are mixed well in the media. DO NOT PIPET.
4. Place the flask with cells in a humidified incubator at 37°C with 5% CO<sub>2</sub>.

### Subculture Procedure

1. Subculture/passage cells when the density reaches 90-100% confluency.
2. Maintain cell density between  $2 \times 10^5$  and  $2 \times 10^6$  viable cells/ml.

### Preparing frozen stocks

*This procedure is designed for 100mm<sup>2</sup> dish or T75cm<sup>2</sup> flask. Scale volumes accordingly to other vessels.*

1. When cells reach 90-100% confluency, freeze down cells.
2. Detach cells according to "Subculture Procedure."
3. Transfer cells to a 15ml conical centrifuge tube and centrifuge at 125 x g for 5 to 7 minutes to collect the cells into a pellet.
4. Carefully aspirate the media and add 1mL of freezing media and gently resuspend by pipetting up and down.
5. Transfer 1mL of cells into a cryogenic vial.
6. Place cryogenic vial in a freezing container (*Nalgene # 5100-0001*) and store at -80°C freezer overnight.
7. Transfer cells to liquid nitrogen for long term storage.